

Review

Unraveling the Mechanisms of Plant Structural Defenses against Insect Pests

¹Aroni Preya Biswas, ¹Roksana Al Nafiu Insha, ¹Md. Khairul Mazed, ¹Md. Mamunur Rahman and ²Md. Motaher Hossain

¹Department of Entomology, Bangabandhu Sheikh Mujibur Rahman Agricultural University, Gazipur, Bangladesh

²Department of Plant Pathology, Bangabandhu Sheikh Mujibur Rahman Agricultural University, Gazipur, Bangladesh

Article history

Received: 29-05-2024

Revised: 03-07-2024

Accepted: 09-08-2024

Corresponding Author:

Md. Mamunur Rahman
Department of Entomology,
Bangabandhu Sheikh Mujibur
Rahman Agricultural
University, Gazipur,
Bangladesh
Email: mamun@bsmrau.edu.bd

Md. Motaher Hossain
Department of Plant
Pathology, Bangabandhu
Sheikh Mujibur Rahman
Agricultural University,
Gazipur, Bangladesh
Email: hossainmm@bsmrau.edu.bd

Abstract: Plants employ a variety of physical, chemical, and genetic strategies to defend against herbivore attacks. These biochemical defense mechanisms are complex and dynamic, encompassing both direct and indirect responses. Defense-related chemicals, whether constitutive or induced by herbivore damage, significantly influence herbivore feeding behavior, growth, and survival. Plant defense systems impact herbivore nutrition, development, and survival, highlighting the importance of chemical substances in reducing plant vulnerability to insects. Plants release volatile chemicals that attract herbivores and these defense strategies are effective alone or in combination. Enhancing our understanding of these defense systems could promote the use of induced resistance as a sustainable pest control strategy, reducing reliance on pesticides. Chemical compounds play a critical role in strengthening plant resistance and understanding the pathways of induced resistance allows for accurate predictions of herbivore responses. Application of induced response elicitors to crops boosts their natural defenses against herbivores. Genetic modification can further enhance plant defenses by promoting the synthesis of defensive chemicals. While this review covers the structural defenses of plants extensively, it lacks a thorough analysis of their ecological impacts beyond deterring insects. Additionally, it focuses more on the mechanisms of these defenses rather than their variations across different ecological contexts or plant species. There is significant potential to leverage plant defense mechanisms in crop protection to address health, environmental, and pesticide-related issues. Utilizing induced resistance, we can develop crop cultivars that naturally respond to minor infections, integrating these traits into sustainable pest management strategies to boost agricultural productivity.

Keywords: Plant Structural Defense, Direct and Indirect Defense, Biotic and Abiotic Stress

Introduction

Plants play a crucial role in maintaining the delicate balance of the Earth's ecosystem and providing energy by warding off diseases and insect herbivores (Musaqaf *et al.*, 2023). Plants have various ways to defend themselves against diseases and pests. Plants have natural defenses to protect themselves from insects and diseases. Plants have developed mechanisms to protect themselves, such as thorns, spines, trichomes, wax coats, and crystal crystals

on their leaves. As noted by Musaqaf *et al.* (2023) in their recent study, plants have developed robust tissues and chemical mechanisms to protect themselves. The productivity of agricultural plants is greatly impacted by the intricate relationship between insect herbivores and plants, as highlighted by Singh and Kaur (2018) in their research. Plant diseases and insect pests present significant challenges to global agricultural productivity. Most crops are susceptible to plant diseases and insect pests. There are various pathogens, including viral,

bacterial, fungal, and nematode. Based on research conducted by Lugtenberg (2016); Dhaliwal *et al.* (2015), it has been found that disease and insect pests have a significant impact on global agricultural output, resulting in losses of 25 and 10.8%, respectively. This category includes harmful insects that feed on foliage or sap and less damaging ones that collect pollen, nectar, or plant resins. Entomologists use the terms "phytophagy" or "phytophagous" to describe these feeding tactics. Grasshoppers and armyworms, being polyphagous insects, possess the remarkable ability to devour the entire plant. However, most insect herbivores tend to be more selective in feeding habits. They have unique abilities that allow them to excel in tasks such as nibbling on leaves, extracting sap, tunneling into stems, pruning roots, creating galls, tunneling through leaves, and gathering pollen or nectar. The symbiotic bond between plants and insects dates back 350 million years. Plants have developed a sophisticated defense mechanism due to their ongoing insect battle. Plants can distinguish between unfamiliar molecules or signals and react by triggering their immune system, much like mammals respond to signals from damaged cells. According to research conducted by Howe and Jander (2008), this process has been observed to activate the plant's natural defense mechanism against herbivores. In nature, various interactions occur, some of which are beneficial, like pollination. However, most interactions involve insects feeding on plants and protecting themselves from these hungry insects. In the world of plants and insects, there is a constant dance between predators and hosts, with every plant species being consumed by at least one insect species. According to a theory put forth by Ehrlich and Raven (1964), the relationship between insects and plants has played a significant role in shaping the diversity of herbivores and their host species. Through co-evolution, both entities have developed strategies to outsmart each other's defensive measures. Howe and Jander (2008) conducted a study that unveiled fascinating plant defense systems that employ morphological and biochemical mechanisms. Hence, it is crucial to tackle plant diseases and insect pests promptly. Traditional pest management methods involve using insecticides and soil amendments to help plants better resist pests (Altieri, 2018). In order to tackle this issue, it might be necessary to use NPK fertilizers and incorporate zinc, boron, magnesium, and silicon. Plants have natural defenses against herbivores or can develop them when attacked. Plants can undergo various changes and produce certain substances when they are attacked by herbivores (Usha Rani and Jyothsna, 2010). Plants utilize various strategies to defend themselves from

herbivores, employing direct and indirect methods. Direct defense alters how organisms choose their host plants and affects their survival and reproductive capabilities. Other species, such as natural enemies of insect pests, offer indirect protection. In 2006, a study was conducted by Dudareva *et al.* Certain plant traits, such as terpenoids, alkaloids, anthocyanins, phenols, and quinones, along with physical defenses like hairs, trichomes, thorns, spines, and thicker leaves, can effectively impede the growth or even cause harm to herbivores (Hanley *et al.*, 2007). Indirect insect defenses are created by producing volatile compounds that lure in predators of herbivores. Enhancing the effectiveness of natural enemies can be achieved by providing them with food and shelter, such as extrafloral nectar (Arimura *et al.*, 2009). Plant defenses against insect herbivores encompass many strategies, including limiting food availability, modifying nutritional content, reducing attractiveness to insects, inflicting physical damage, and hindering chemical processes. Various compounds produced by plants play a vital role in their defense mechanisms, acting as important chemical defenses against insects and other threats. Understanding the interactions between herbivores and plants is paramount and necessitates a multidisciplinary approach within plant biology. Understanding the chemical and ecological factors that influence these relationships requires the integration of multiple disciplines. Our current knowledge of chemical interactions between plants and other organisms, including neighbors, symbionts, diseases, herbivores, and natural enemies, is still limited. This fascinating region shows great potential for crop protection. Understanding the genetic composition of plant defense mechanisms can greatly contribute to creating crops with natural resistance against herbivores.

Therefore, reducing reliance on harmful insecticides is imperative for effective insect control. The ongoing evolutionary conflict between plants and herbivores suggests potential co-evolution, where herbivores adapt to resistant plant genotypes. Understanding the intricate relationships between plants and herbivores is crucial for enhancing crop productivity. This review comprehensively explores diverse plant defense mechanisms employed against insect pests. It aims to provide a detailed account of both direct and indirect defense strategies plants use to fend off herbivores. Moreover, the study seeks to offer practical insights into leveraging these mechanisms to optimize agricultural practices. This approach underscores the importance of a holistic understanding and application of plant defenses in integrated pest management strategies.

Materials and Methods

This study is entirely a review, relying exclusively on secondary sources, including relevant books, e-journals, research articles, scientific reports, and bulletins. Internet browsing was also used to make sure the data was up to date. Following the collection of all relevant data, it was methodically gathered and arranged chronologically to support the goals of this study.

Results and Discussions

Direct Defenses

Plants have developed various ways to protect themselves against herbivores. One of these is through their structural features, like the thickness and lignification of cell walls, thorns, trichomes, and the presence of wax on leaf surfaces. Furthermore, secondary metabolites have the potential to act as toxins, impacting the growth, development, and digestibility of herbivores. According to a study by Agrawal *et al.* (2009), this process creates barriers that effectively safeguard the plant against future attacks. Furthermore, the combined efforts of different protective factors strengthen the plant's capacity to repel herbivorous intruders. When tomatoes are consumed individually, the effects of alkaloids, phenolics, Proteinase Inhibitors (PIs), and oxidative enzymes on insects are diminished. However, the interaction between these substances has a powerful impact on the insect, influencing it in various ways such as ingestion, digestion, and metabolism. Through the collaboration of trypsin proteinase inhibitors and nicotine production, *Nicotiana attenuata* enhances its defense against the insect *Spodoptera exigua*.

Morphological Features for Physical Defense

Understanding plant structures is crucial for safeguarding plants against herbivory and plays a vital role in the defense mechanisms of host plants against insects, commonly referred to as Host Plant Resistance (HPR). Plants have evolved a multitude of strategies to protect themselves from insect pests. One approach is to establish a physical barrier, which can be achieved by developing a waxy cuticle or by growing spines, setae, and trichomes. Structural defenses are the physical characteristics of plants that provide them with an advantage by discouraging herbivores from feeding. Plants have a wide array of defenses, which can range from easily observable characteristics to more subtle changes in cell wall thickness caused by lignification and suberization. Plants protect themselves against herbivory in large part through the incorporation of minerals into their tissues, the presence of spines and thorns, hair-like structures on their leaves, toughened or hardened leaves, and a branching pattern that produces shoots at wide angles from the main stem (He *et al.*, 2011; Chamarthi *et al.*, 2011). Spinescence

encompasses a range of botanical features such as spines, thorns, and prickles. A study conducted by Hanley *et al.* (2007) has revealed that this specific plant has innate mechanisms to protect itself from various insects. During pubescence, plants develop small hair-like structures called trichomes on different parts of their bodies, including the stem, leaves, and fruits. The trichomes can exhibit different shapes, such as straight, spiral, stellate, hooked, and glandular, as noted by Hanley *et al.* (2007). Chamarthi *et al.* (2011) conducted research and discovered that the resistance of sorghum *Sorghum bicolor* (L.) to the shoot fly *Atherigona toccata* (Rondani) is associated with the shiny look of the leaves, as well as the coloration of the plumule and leaf sheath. The role of various morphological features in defending plants against insect pests is summarized in Table (1).

Table 1: The role of plant morphology in defending against insect pests

Features	Function	References
Surface waxes	<ul style="list-style-type: none"> •The presence of waxes on the surface of plants allows insects to detect harmful chemicals and physical sensations, which in turn helps the plant defend itself against insect attacks 	Blenn <i>et al.</i> (2012)
Trichomes	<ul style="list-style-type: none"> •Have an impact on pest herbivorous insects' search patterns •Exploring the various morphological adaptations of plants to counter insect pests 	Yang <i>et al.</i> (2023); Howe and Schaller (2008)
Thickening of cell walls	<ul style="list-style-type: none"> •Plant cell walls undergo a process that makes them rigid and durable, which in turn makes them less susceptible to damage caused by insect pests, such as tearing by their jaws or penetration by their egg-laying organs 	Raup (1985)
Plant color	<ul style="list-style-type: none"> •Adult <i>Pieris rapae</i> has a preference for green and blue-green surfaces while engaging in preovipositional displays •The cabbage aphid <i>Brevicoryne brassicae</i> has reduced attraction towards red Cruciferae 	Lev-Yadun (2016)

Accumulation of minerals in plant cuticle	•Accumulation of Silicon (Si) can provide plants with a defense mechanism against herbivores who gnaw on them	Islam <i>et al.</i> (2022)
---	---	----------------------------

Direct Chemical Defense

Plant direct defense measures to prevent insect pest infestation provides an overview of the major chemical defenses' plants use against insect pests, categorized by their mode of action and specific effects on pest physiology or behavior (Table 2).

Constitutive Defenses vs Induced Defenses

Two main strategies used by plants to fend off insect pests are constitutive and induced defenses. Physical barriers like trichomes and cuticles, as well as chemical defenses like secondary metabolites, are examples of constitutive defenses, which are always present (Howe and Jander, 2008). Regardless of the presence of pests, these protections serve as a baseline defensive system, offering consistent resistance. On the other hand, induced defenses are triggered in reaction to environmental stressors or herbivore attacks. To draw in natural adversaries, these defenses may include the release of volatile organic compounds, the synthesis of protective proteins, or the overexpression of particular secondary metabolites (Karban and Baldwin, 2007). Induced defenses are more resource-efficient, deploying only, when necessary, whereas constitutive defenses are energy-intensive and maintained regardless of threats. This reflects an adaptive strategy to balance defense and growth (Züst and Agrawal, 2017). The different tactics that plants employ to defend themselves against herbivores are highlighted in Fig. (1), which distinguishes between constitutive and induced defenses.

Silicon-Mediated Plant Defense against Pathogens and Insect Pests

Recent studies have highlighted the importance of this research in addressing both biological and environmental challenges in plants, as shown by the findings of Leroy *et al.* (2019); Seal *et al.* (2018); and Zargar *et al.* (2019). Studies have demonstrated that the use of silicon can boost the ability of important crops to withstand different diseases and pests (Liang *et al.*, 2015; Song and Thomma, 2018). Studies have indicated that silicon plays a crucial role in essential metabolic processes, as demonstrated by the research conducted by Frew *et al.* (2018); and Coskun *et al.* (2019). Table (3) presents the silicon-mediated defense mechanism against insect pest attacks in plants. According to a recent study, researchers examined how Si affects the growth of *Saccharum* spp. Hybrids and their ability to defend against insect herbivores. The study found noteworthy

enhancements in plant growth and productivity (Frew *et al.*, 2018). According to a study conducted by Chain *et al.* (2009), the utilization of silicon had a noteworthy influence on the regulation of 47 genes in *Triticum aestivum* plants. In a study conducted by Brunings *et al.* (2009), it was found that the use of silicon on ordinary rice plants had a significant impact on gene regulation. Specifically, 221 genes were affected, with 28 of them being linked to defense and stress responses.

Table 2: Plant direct defense measures to prevent insect pest infestation

Features	Functions	References
Bioactive specialized compounds	Primary metabolites play a crucial role in supporting growth, development, and reproduction. Plants have the ability to produce secondary metabolites like alkaloids, terpenoids, and phenolics as a defense mechanism against insect herbivores	Matsui <i>et al.</i> (2022)
Hypersensitive activity	The intruder causes the growth of necrotic plant tissue, which separates it from the plants and lays eggs on their leaves	Jeblick <i>et al.</i> (2023)
Digestibility reduction	The presence of defensive proteins, such as protein inhibitors, lectin, chitinase, and polyphenol oxidase, decreases the activity of insect herbivores in digesting the plant	Silva-Junior <i>et al.</i> (2022)

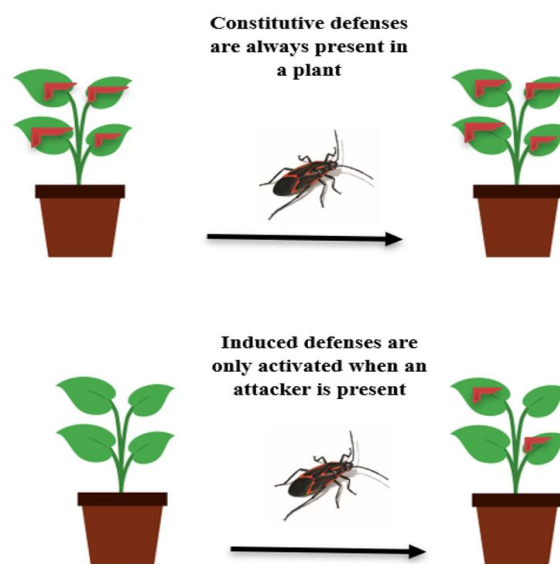


Fig. 1: Constitutive defenses vs induced defenses

Table 3: Silicon-mediated defense mechanism against insect pest attack in plants

Defense mechanism	Description	Reference
Silicon deposition	Plants accumulate silicon in their tissues, forming a physical barrier against insect penetration	Ma and Yamaji (2006)
Induced resistance	Silicon enhances plant resistance by inducing Systemic Acquired Resistance (SAR) against pests	Keeping <i>et al.</i> (2009)
Increased lignification	Silicon application leads to increased lignin deposition, strengthening cell walls against insect-feeding	Reynolds <i>et al.</i> (2016)
Alteration of insect behavior	Silicon-treated plants alter insect feeding behavior, reducing pest damage	Massey <i>et al.</i> (2006)
Regulation of plant hormones	Silicon modulates the expression of plant hormones involved in defense responses against insects	Fauteux <i>et al.</i> (2005)

Biochemical Defense Mechanisms

Alkaloids

Alkaloids are a diverse group of biochemicals that are present in a variety of living organisms, with plants being the main origin (Levin, 1973). These compounds can be found in a wide range of plant species, such as Leguminosae, Liliaceae, Solanaceae, and Amaryllidaceae. It is thought that they may have developed as a means of protecting themselves from insect herbivory (Howe and Jander, 2008). Alkaloids are produced from amino acids in the roots and then transported through the phloem and sometimes the xylem, eventually building up above ground (Courdavault *et al.*, 2014). Alternatively, the final stages of their de novo biosynthesis can take place above the surface, as mentioned in a study by Miettinen *et al.* (2014). It's interesting to observe that certain plants produce nectar with trace amounts of alkaloids. This serves a dual purpose: It functions as a defense mechanism against nectar robbers and impacts the behavior of their natural pollinators, thus increasing their reproductive success. These substances contain nitrogen within a heterocyclic ring structure. The ring structure is composed of a wide range of compounds, including pyridines, pyrroles, indoles, pyrrolidines, isoquinolines, and piperidines (Berlinck and Kossuga, 2007).

Plant Defensive Proteins

It has been discovered through recent advancements in microarray and proteomic approaches that there is a diverse range of plant defense mechanisms against herbivores. Various signaling pathways, such as Jasmonic Acid (JA), SA and Ethylene (ET), play a role in regulating arthropod-inducible proteins due to the wide range of feeding habits exhibited by arthropods (Chen *et al.*, 2005). Table (4) summarizes the plant defensive proteins against insect pests, providing insights into the specific proteins involved in these defense strategies.

Indirect Defenses

Understanding the defensive response in plants and how it attracts natural enemies of herbivores is crucial for protecting plants from herbivore attacks (Dudareva *et al.*, 2006). Indirect defenses can be either constitutive or induced due to the combined effects of mechanical damage and elicitors from the herbivore. The production of volatiles and the secretion of Extra Floral Nectar (EFN) play a crucial role in how plants interact with natural enemies of insect pests, such as parasitoids or predators. These interactions actively work to decrease the population of feeding herbivores (Dudareva *et al.*, 2006). Induced indirect defenses have been the subject of growing interest in recent times and have been extensively researched across various disciplines, including genetics, biochemistry, physiology, and ecology (Maffei *et al.*, 2010).

Role of Phytohormones in Induced Resistance in Plants

Recognizing plant defense against herbivore attack requires a deep understanding of the intricate signal transduction pathways and the complex network of phytohormones involved. Understanding the intricate workings of plant hormones is essential for comprehending how plants grow, develop, and protect themselves.

Table 4: Plant defensive proteins against insect pests

Putative defense protein	Plant species	Insect species	Reference
PIs	<i>Sorghum bicolor</i>	<i>Schizaphis graminum</i>	Zhu-Salzman <i>et al.</i> (2004); Dunse <i>et al.</i> (2010); Hartl <i>et al.</i> (2011);
	Tomato	<i>Manduca sexta</i>	Steppuhn and Baldwin (2007)
	<i>Solanum nigrum</i>	<i>Manduca sexta</i>	
	<i>Nicotiana attenuata</i>	<i>Spodoptera littoralis</i>	
	Transgenic Arabidopsis/oil seed rape	<i>Spodoptera exigua</i>	
		<i>Spodoptera exigua</i>	
		<i>Spodoptera exigua</i>	
LOXs	<i>Nicotiana attenuata</i>	<i>Bemisia tabaci</i>	Kempema <i>et al.</i> (2007)
	Wheat	<i>Sitobion avenae</i>	
	<i>Nicotiana attenuata</i>	<i>Myzus nicotianae</i>	
Chitinases	<i>Sorghum bicolor</i>	<i>Schizaphis graminum</i>	Zhu-Salzman <i>et al.</i> (2004)

Role of Phenolic Compounds in Plant-Defensive Mechanisms

Phenolic compounds are secondary metabolites that can be found in plants across the board. These pathways, as described in previous studies (Balasundram *et al.*, 2006; Cheynier, 2012), give rise to the derivatives of pentose phosphate, shikimate, and phenylpropanoid. Tables (5-6) describe how phytohormones contribute to the development of plant resistance and show the critical role that certain naturally occurring phenolic compounds play in these defense mechanisms. Additionally in Fig. (2), how plant secondary metabolites playing role against herbivores is discussed.

Table 5: The function of phytohormones in the process of induced resistance in plants

Phytohormones names	Function
Jasmonic acid	<ul style="list-style-type: none"> Induces the activation of both direct and indirect defensive mechanisms (War <i>et al.</i>, 2011) Numerous genes responsible for herbivore defense are controlled by JA, as demonstrated by Shivaji <i>et al.</i> (2010)
Salicylic acid	<ul style="list-style-type: none"> A crucial phytohormone that induces many metabolic and physiological reactions in plants, especially in defense mechanisms Rivas-San Vicente <i>et al.</i> (2011)
Ethylene	<ul style="list-style-type: none"> Plant defense against herbivores and pathogens can be triggered both directly and indirectly van Loon <i>et al.</i> (2006) The presence of <i>A. alni</i> caused the release of ethylene Tschardt <i>et al.</i> (2001)

Table 6: Role of some naturally-occurring phenolic compounds

Phenolic compound	Function	Reference
Flavonoids	Antioxidant, anti-inflammatory	Middleton <i>et al.</i> (2000)
Catechins	Antioxidant, anti-inflammatory	Arts and Hollman (2005)
Resveratrol	Antioxidant, cardio-protective	Baur and Sinclair (2006)
Quercetin	Anti-inflammatory, immune support	Boots <i>et al.</i> (2008)
Ellagic Acid	Anticancer, antimicrobial	González-Sarrías <i>et al.</i> (2017)
Curcumin	Anti-inflammatory, antioxidant	Aggarwal and Harikumar (2009)

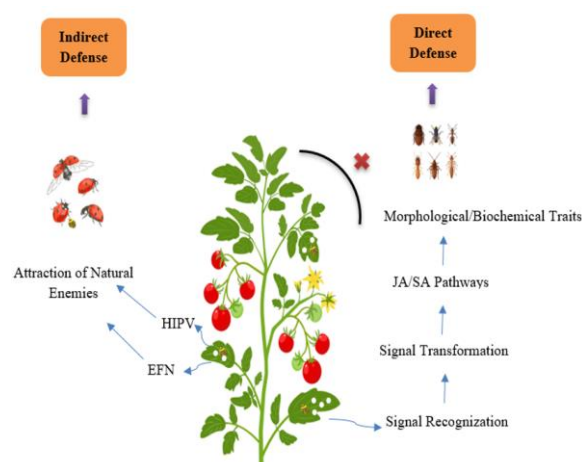


Fig. 2: Plant secondary metabolites as defense tools against herbivores

Plant Secondary Metabolites as Defense Tools against Herbivores for Sustainable Crop Protection

PSMs are biologically active substances that have the ability to repel or intoxicate insects and disrupt their digestion. Alkaloids have been found to be unappealing to insect pests, serving as deterrents to their feeding and inhibiting their growth. They also target neurotransmitters, disrupting neuronal signal transduction (Pavela *et al.*, 2016). Alkaloids have a negative impact on the concentrations and expression of neurotransmitters. These changes have a negative impact on the physiology and behavior of insects, resulting in direct toxicity or a lack of preference for the host they feed on. Some examples of alkaloids that have an impact on neuronal signal transduction include nicotine, caffeine, erythrina alkaloids, tubocurarine, ergot alkaloids, muscarine, agroclavine, and theophylline. Research has found that caffeine, an alkaloid, has demonstrated insecticidal properties. It has been observed to cause paralysis and intoxication in herbivores by inhibiting phosphodiesterase activity. This discovery suggests that caffeine could be a promising option for use as a biopesticide (Hollingsworth *et al.*, 2002). It was discovered that nicotine, an alkaloid found in tobacco, was unintentionally employed for controlling insect pests (Pavela *et al.*, 2016). Various secondary metabolites, like pyrethrum from certain plants in the Asteraceae family, azadirachtin derived from neem seeds, and capsaicin obtained from hot pepper extracts, have been utilized as insecticides (Benelli *et al.*, 2017). These metabolites have various effects on insects, such as blocking receptors and channels in their nervous system, inhibiting cellular respiration, and disrupting their hormonal balance (Rattan *et al.*, 2010). Here, the adverse effects of plant secondary metabolites that activate defense mechanisms against insect pests are discussed in Table (7).

Table 7: Adverse effect of plant secondary metabolites that activate in defense mechanism against insect pest

Adverse effect	Description	Reference
Reduced growth and development	PSMs can inhibit nutrient absorption and metabolism, leading to slower growth rates and developmental delays in herbivore insects	Figuroa-Macias <i>et al.</i> (2021)
Altered feeding behavior	PSMs may deter herbivores through aversion or reduced feeding rates due to unpleasant taste or toxicity	Bernays and Chapman (2007)
Disruption of reproduction	Some PSMs can interfere with reproductive processes in herbivore insects, leading to decreased fecundity or viability of offspring	Raubenheimer <i>et al.</i> (2009)
Increased mortality	Toxic PSMs can cause direct mortality in herbivore insects by poisoning or disrupting vital physiological functions	Petschenka and Agrawal (2016)
Alteration of developmental timing	PSMs may disrupt normal developmental sequences in herbivore insects, resulting in altered phenotypes or life cycle durations	Schoonhoven <i>et al.</i> (2005)
Induction of Behavioral Avoidance Strategies	Herbivore insects may evolve behavioral mechanisms to avoid plants containing high levels of PSMs, reducing their fitness	Dethier (1976)

Ecological Impacts of Plant Structural Defenses on Insect Populations and Communities

Plant structural defenses play pivotal roles in shaping insect populations, behaviors, community structures, and evolutionary dynamics. Firstly, these defenses directly influence insect populations by physically deterring herbivores. For instance, the hardness of leaves can decrease the feeding efficiency of herbivorous insects, thereby reducing their rates of development and reproductive success (Carmona *et al.*, 2011). Plants with tougher leaves tend to sustain lower levels of herbivore damage and support smaller populations of generalist herbivores compared to those with more delicate foliage.

Additionally, trichomes, such as those found on tomato plants, effectively hinder insect movement and feeding, leading to significant reductions in pest populations like whiteflies (Watson *et al.*, 2015).

Plant structural defenses alter insect behavior by impeding access to and consumption of plant tissues. Insects confronted with these defenses often expend more energy and time overcoming them, resulting in reduced feeding efficiency and overall fitness. For example, glandular trichomes on cotton plants induce heightened grooming behavior in larvae of *Helicoverpa armigera*, which subsequently decreases their feeding activity and growth rates (Amin *et al.*, 2014). This behavioral adaptation not only highlights the potential for managing herbivorous insect populations but also underscores the ecological implications of plant defenses in agricultural and natural ecosystems.

Structural defenses exert profound impacts on insect community structures. By selectively deterring specific herbivores, these mechanisms create specialized habitats that can foster greater biodiversity within insect communities. Furthermore, plants equipped with robust structural defenses attract distinct groups of herbivores and their predators, thereby influencing community dynamics. For instance, the presence of spines on certain plant species in tropical forests correlates with increased diversity among plant-eating insects and their predators, enriching the complexity of insect communities and demonstrating the ecological significance of plant defenses (Glassmire *et al.*, 2016). These interactions underscore the dynamic relationships between plants and insects and highlight the evolutionary implications of co-adaptation and competition between herbivores and their plant hosts (Ferlian *et al.*, 2020).

Co-Evolution between Plants and Insect Pests

The co-evolutionary relationship between plants and insect pests is a dynamic and ongoing process characterized by reciprocal selective pressures that drive continuous adaptation and counter-adaptation. This interaction profoundly influences the evolution of both parties, shaping their genetic makeup, behaviors and ecological interactions.

Insects, exemplified by species like the Colorado potato beetle, demonstrate rapid evolution of resistance to plant defenses. This scenario illustrates a classic co-evolutionary arms race where plants develop new chemical defenses and pests swiftly evolve mechanisms to overcome these defenses. For instance, beetle populations can acquire resistance within a few generations, underscoring the adaptive capacity of insect pests under selective pressure (Griese, 2021).

Host shifts among insect pests, such as the case of the apple maggot fly (*Rhagoletis pomonella*), can lead to significant evolutionary consequences. The transition

from hawthorn to apple as a preferred host has driven genetic divergence and early stages of speciation within the fly population. This adaptation highlights how changes in host preferences can drive diversification in both plants and insects, influencing their ecological and evolutionary trajectories (Farley-Barnes *et al.*, 2019).

Plants have evolved diverse chemical defenses to deter herbivores, leading to intricate co-evolutionary dynamics. For example, the development of glucosinolates in the Brassicaceae family has reduced damage from generalist herbivores while simultaneously increasing vulnerability to specialized pests like the cabbage white butterfly (*Pieris rapae*). This selective pressure drives adaptations in pests to detoxify or bypass these chemical defenses, illustrating a co-evolutionary arms race in biochemical interactions (Nallu *et al.*, 2018).

In response to plant defenses, certain insect pests have evolved specific morphological adaptations. For instance, beetles have developed enlarged mouthparts to circumvent physical barriers and reach protected plant tissues. Similarly, plants may undergo morphological changes such as leaf thickening or the formation of physical barriers in response to these adaptations, illustrating the reciprocal morphological evolution driven by co-evolutionary pressures (Arora and Sandhu, 2017).

Advancements in genomic technologies provide valuable insights into the genetic basis of co-evolutionary interactions between plants and insect pests. Studies on organisms like the diamondback moth (*Plutella xylostella*) have identified specific genes associated with resistance to plant toxins, revealing the intricate genomic adaptations that underpin the evolutionary arms race between plants and herbivores. These genomic insights highlight the role of genetic variation in driving adaptation and counter-adaptation in co-evolutionary processes (Ma *et al.*, 2021).

Adaptation and Diversification of Defense Strategies in Different Plant Species

Plants have evolved a diverse array of defense strategies in response to ecological pressures, demonstrating remarkable adaptation and diversification to ward off herbivores. These strategies encompass chemical defenses, where plants synthesize toxic or repellent compounds like glucosinolates in the Brassicaceae family, which are activated upon tissue damage to deter herbivory (Chhajed *et al.*, 2020). Additionally, structural defenses such as thorns and trichomes physically obstruct herbivores and can foster mutualistic relationships, as seen in Acacia plants that host defensive ants (Schulze *et al.*, 2019).

Indirect defenses involve plants releasing Volatile Organic Compounds (VOCs) in response to herbivore attacks, which attract natural predators like parasitoid wasps. This strategy exemplifies how plants use

chemical signaling to enlist the help of predators, thereby reducing herbivore populations indirectly (Turlings and Erb, 2018). Furthermore, tolerance strategies mitigate herbivory's impact on plant fitness by promoting rapid regrowth or by synchronizing life cycle events like flowering time to avoid peak herbivore activity, as observed in grasses and alpine plants, respectively (Forrister, 2022; Silva *et al.*, 2021).

Genetic diversification within plant populations contributes significantly to defense strategy evolution. Species like *Solanum peruvianum* exhibit substantial genetic diversity in their defenses against herbivores, allowing them to adapt to local herbivore pressures and environmental conditions (Tan, 2019). This variability underscores the role of genetic adaptation in shaping plant defense mechanisms and enhancing their resilience in diverse ecological contexts.

Applications in Agriculture and Pest Management

Tomato plants (*Solanum lycopersicum*) include glandular trichomes that release a range of substances, including acylsugars and volatile chemicals, that discourage herbivorous insects like whiteflies and spider mites (Naalden *et al.*, 2021). Integrated pest management solutions utilize trichome-mediated defenses by choosing cultivars with a high density of trichomes to naturally decrease insect infestations (Glas *et al.*, 2012).

Rice plants (*Oryza sativa*) have the ability to store silica in their tissues. This accumulation of silica strengthens the plant's structural defenses against insect pests such as rice stem borers and leaf folder caterpillars (Rosenhek-Goldian *et al.*, 2015). Integrated pest management utilizes the buildup of silica by encouraging farming practices that increase the availability of silicon in rice fields. This helps to decrease insect damage and the need for pesticides (Curto *et al.*, 2016).

Coniferous trees, such as pine (*Pinus spp.*) and spruce (*Picea spp.*), possess resin ducts that carry poisonous secondary metabolites such as terpenes and phenolics. These substances act as deterrents against bark beetles and other insects that bore into wood (Paljakka, 2020). Integrated pest management tactics leverage resin-based defenses by advocating for forest management approaches that sustain robust resin production in trees, hence decreasing vulnerability to insect outbreaks (Whitehill *et al.*, 2023).

Recent Case Studies on Plant Defense Mechanisms Against Herbivores

Plants have evolved diverse and sophisticated defense mechanisms to protect themselves from herbivorous insects, showcasing remarkable adaptations across different species. Trichomes, for example, found on tomato leaves physically impede insect movement and reduce feeding efficiency, thereby decreasing damage

from pests such as aphids and spider mites (Aljbory and Chen, 2018). Similarly, grasses like rice and wheat accumulate silica in their tissues, enhancing mechanical strength and resilience against chewing insects like stem borers and leafhoppers (Greenslade *et al.*, 2016).

Another significant defense strategy involves the production of leaf waxes and cuticular layers rich in compounds that deter insect feeding and oviposition, as observed in Arabidopsis plants. Genetic studies have highlighted that mutants lacking effective wax biosynthesis pathways show increased susceptibility to herbivorous insects, underscoring the role of these cuticular defenses (Hehmeyer, 2019). Furthermore, Acacia trees develop extrafloral nectaries that attract ants, which act as defenders against herbivores through predation or deterrence. Field experiments have demonstrated that ant-tended Acacia plants experience reduced herbivory compared to untended plants, highlighting the effectiveness of indirect defenses (Dejean *et al.*, 2013).

In addition to chemical and indirect defenses, morphological adaptations play a crucial role in plant defense strategies. Cacti, with their succulent stems and reduced leaves, minimize water loss and reduce attractiveness to herbivorous insects. Comparative studies across cactus species reveal that succulent varieties experience lower herbivory due to their reduced leaf area and water content (Hunter *et al.*, 2020). Eucalyptus trees exhibit another morphological adaptation with thick bark layers that provide physical protection against both herbivorous insects and environmental stressors like fire. Research shows that species with thicker bark layers suffer less damage from insects and demonstrate greater resilience in challenging environmental conditions (Lindenmayer and Franklin, 2013).

These case studies underscore the diverse and adaptive nature of plant defense mechanisms against herbivores, ranging from chemical deterrents and physical barriers to morphological adaptations and symbiotic relationships with other organisms. Understanding these mechanisms not only enhances our appreciation of plant resilience but also informs strategies for sustainable pest management in agriculture and natural ecosystems.

Limitations of the Study

This review article on plant defense mechanisms against insect herbivores provides a comprehensive overview but is subject to several limitations. While it covers a wide range of structural and chemical defenses employed by plants, the depth of analysis into specific molecular pathways and emerging research areas could be enhanced. There is a temporal bias as it primarily synthesizes studies up to recent years, potentially

overlooking newer developments and regional variations in defense strategies. The focus on agricultural contexts might limit insights into natural ecosystems, where different ecological dynamics may influence plant-insect interactions differently. Generalizations across diverse plant species and insect communities could oversimplify the variability in defense responses. Moreover, reliance on published studies may introduce biases and overlook unpublished or negative findings. To address these limitations, future reviews could aim for a more nuanced examination of molecular mechanisms, incorporate emerging research trends, explore non-agricultural settings, and consider interdisciplinary perspectives to deepen understanding. Additionally, ensuring the representation of diverse experimental conditions and acknowledging practical challenges in implementing plant defense strategies at scale would enhance the relevance and applicability of such reviews in agricultural and ecological management.

Conclusion

The findings have explored the intriguing realm of plant defense systems against insect herbivores. By studying the morphological, biochemical, and molecular approaches, we have acquired knowledge of the extraordinary methods by which plants have adapted to defend themselves against herbivores. Gaining knowledge of these systems is not only scientifically intriguing but also has substantial practical consequences for pest control in agriculture. This highlights the significance of utilizing plant defenses in integrated pest management tactics, ultimately aiding in developing more sustainable and resilient agricultural practices. Plant defenses impede herbivores' ability to get nutrients. Reduced plant fitness occurs when herbivore stress and weaker defenses increase herbivore damage. Plants do not possess nervous systems, yet they utilize signaling pathways to carry out defense mechanisms that are direct (such as poisons and reallocating resources) and indirect (such as attracting natural enemies). Utilizing these processes is crucial for managing insect pests and implementing integrated pest control in agriculture.

Acknowledgment

The authors acknowledge the Research Management Wing (RMW) of Bangabandhu Sheikh Mujibur Rahman Agricultural University, Bangladesh for providing the support for writing this manuscript.

Funding Information

The authors have not received any financial support or funding to report.

Author's Contributions

Aroni Preya Biswas: Coordinated data collection, performed data analysis and contributed to manuscript writing.

Roksana Al Nafiu Insha: Interpreted results and contributed to manuscript writing.

Md. Khairul Mazed: Verified results and contributed to the discussion section of the manuscript.

Md. Mamunur Rahman: Conceived and designed the research, implemented the study, analyzed the results, and contributed to manuscript writing.

Md. Motaher Hossain: Interpreted results and contributed to manuscript writing.

Ethics

This review article is original and contains unpublished material. The corresponding authors confirm that other authors have read and approved the manuscript and no ethical issues involved.

Conflict of Interest

The authors have no conflict of interest.

References

- Agrawal, A. A., Fishbein, M., Jetter, R., Salminen, J., Goldstein, J. B., Freitag, A. E., & Sparks, J. P. (2009). Phylogenetic ecology of leaf surface traits in the milkweeds (*Asclepias* spp.): chemistry, ecophysiology, and insect behavior. *New Phytologist*, 183(3), 848–867. <https://doi.org/10.1111/j.1469-8137.2009.02897.x>
- Aljbory, Z., & Chen, M. (2018). Indirect plant defense against insect herbivores: a review. *Insect Science*, 25(1), 2–23. <https://doi.org/10.1111/1744-7917.12436>
- Altieri, M. A. (2018). *Agroecology: the science of sustainable agriculture*. CrC Press.
- Amin, Md. R., Azad, H. M. S., Hossain, Md. Shamim., Suh, S. J., & Kwon, Y. J. (2014). Foraging Behavior of *Helicoverpa armigera* Hübner (*Lepidoptera: Noctuidae*) First Instar Larvae on Selected Cotton Varieties. *Current Research on Agriculture and Life Sciences*, 32(4), 185–188. <https://doi.org/10.14518/crals.2014.32.4.033>
- Arimura, G., Matsui, K., & Takabayashi, J. T. (2009). Chemical and Molecular Ecology of Herbivore-Induced Plant Volatiles: Proximate Factors and Their Ultimate Functions. *Plant and Cell Physiology*, 50(5), 911–923. <https://doi.org/10.1093/pcp/pcp030>
- Arora, R., & Sandhu, S. (2017). Insect-Plant Interrelationships. *Breeding Insect Resistant Crops for Sustainable Agriculture*, 1–44. https://doi.org/10.1007/978-981-10-6056-4_1
- Arts, I. C., & Hollman, P. C. (2005). Polyphenols and disease risk in epidemiologic studies. *The American Journal of Clinical Nutrition*, 81(1), 317S–325S. <https://doi.org/10.1093/ajcn/81.1.317s>
- Balasundram, N., Sundram, K., & Samman, S. (2006). Phenolic compounds in plants and agri-industrial by-products: Antioxidant activity, occurrence, and potential uses. *Food Chemistry*, 99(1), 191–203. <https://doi.org/10.1016/j.foodchem.2005.07.042>
- Baur, J. A., & Sinclair, D. A. (2006). Therapeutic potential of resveratrol: The in vivo evidence. *Nature Reviews Drug Discovery*, 5(6), 493–506. <https://doi.org/10.1038/nrd2060>
- Benelli, G., Canale, A., Toniolo, C., Higuchi, A., Murugan, K., Pavela, R., & Nicoletti, M. (2017). Neem (*Azadirachta indica*): towards the ideal insecticide? *Natural Product Research*, 31(4), 369–386. <https://doi.org/10.1080/14786419.2016.1214834>
- Berlinck, R., & Kossuga, M. (2007). *Modern Alkaloids: Structure, Isolation, Synthesis and Biology*. <https://doi.org/10.1002/9783527621071.ch11>
- Bernays, E. A., & Chapman, R. F. (2007). *Host-plant selection by phytophagous insects*. 2.
- Blenn, B., Bandoly, M., Küffner, A., Otte, T., Geiselhardt, S., Fatouros, N. E., & Hilker, M. (2012). Insect Egg Deposition Induces Indirect Defense and Epicuticular Wax Changes in *Arabidopsis thaliana*. *Journal of Chemical Ecology*, 38(7), 882–892. <https://doi.org/10.1007/s10886-012-0132-8>
- Boots, A. W., Haenen, G. R. M. M., & Bast, A. (2008). Health effects of quercetin: From antioxidant to nutraceutical. *European Journal of Pharmacology*, 585(2–3), 325–337. <https://doi.org/10.1016/j.ejphar.2008.03.008>
- Brunings, A. M., Datnoff, L. E., Ma, J. F., Mitani, N., Nagamura, Y., Rathinasabapathi, B., & Kirst, M. (2009). Differential gene expression of rice in response to silicon and rice blast fungus *Magnaporthe oryzae*. *Annals of Applied Biology*, 155(2), 161–170. <https://doi.org/10.1111/j.1744-7348.2009.00347.x>
- Carmona, D., Lajeunesse, M. J., & Johnson, M. T. J. (2011). Plant traits that predict resistance to herbivores. *Functional Ecology*, 25(2), 358–367. <https://doi.org/10.1111/j.1365-2435.2010.01794.x>
- Chain, F., Côté-Beaulieu, C., Belzile, F., Menzies, J. G., & Bélanger, R. R. (2009). A Comprehensive Transcriptomic Analysis of the Effect of Silicon on Wheat Plants Under Control and Pathogen Stress Conditions. *Molecular Plant-Microbe Interactions®*, 22(11), 1323–1330. <https://doi.org/10.1094/mpmi-22-11-1323>

- Chamarthi, S. K., Sharma, H. C., Sahrawat, K. L., Narasu, L. M., & Dhillon, M. K. (2011). Physico-chemical mechanisms of resistance to shoot fly, *Atherigona soccata* in sorghum, *Sorghum bicolor*. *Journal of Applied Entomology*, 135(6), 446–455. <https://doi.org/10.1111/j.1439-0418.2010.01564.x>
- Chen, H., Wilkerson, C. G., Kuchar, J. A., Phinney, B. S., & Howe, G. A. (2005). Jasmonate-inducible plant enzymes degrade essential amino acids in the herbivore midgut. *Proceedings of the National Academy of Sciences*, 102(52), 19237–19242. <https://doi.org/10.1073/pnas.0509026102>
- Cheyrier, V. (2012). Phenolic compounds: from plants to foods. *Phytochemistry Reviews*, 11(2–3), 153–177. <https://doi.org/10.1007/s11101-012-9242-8>
- Chhajed, S., Mostafa, I., He, Y., Abou-Hashem, M., El-Domiaty, M., & Chen, S. (2020). Glucosinolate Biosynthesis and the Glucosinolate–Myrosinase System in Plant Defense. *Agronomy*, 10(11), 1786. <https://doi.org/10.3390/agronomy10111786>
- Coskun, D., Deshmukh, R., Sonah, H., Menzies, J. G., Reynolds, O., Ma, J. F., Kronzucker, H. J., & Bélanger, R. R. (2019). The controversies of silicon's role in plant biology. *New Phytologist*, 221(1), 67–85. <https://doi.org/10.1111/nph.15343>
- Courdavault, V., Papon, N., Clastre, M., Giglioli-Guivarc'h, N., St-Pierre, B., & Burlat, V. (2014). A look inside an alkaloid multisite plant: the *Catharanthus* logistics. *Current Opinion in Plant Biology*, 19, 43–50. <https://doi.org/10.1016/j.pbi.2014.03.010>
- Curto, G., Dallavalle, E., Matteo, R., & Lazzeri, L. (2016). Biofumigant effect of new defatted seed meals against the southern root-knot nematode, *Meloidogyne incognita* *Annals of Applied Biology*, 169(1), 17–26. <https://doi.org/10.1111/aab.12275>
- Dejean, A., Orivel, J., Rossi, V., Roux, O., Lauth, J., Malé, P.-J. G., Céréghino, R., & Leroy, C. (2013). Predation Success By A Plant-Ant Indirectly Favours The Growth And Fitness Of Its Host Myrmecophyte. *PLoS ONE*, 8(3), e59405. <https://doi.org/10.1371/journal.pone.0059405>
- Dethier, V. G. (1976). *The hungry fly: a physiological study of the behavior associated with feeding*.
- Dhaliwal, G. S., Jindal, V., & Mohindru, B. (2015). Crop Losses due to insect pests: Global and Indian Scenario. *Indian Journal of Entomology*, 77(2), 165. <https://doi.org/10.5958/0974-8172.2015.00033.4>
- Dudareva, N., Negre, F., Nagegowda, D. A., & Orlova, I. (2006). Plant Volatiles: Recent Advances and Future Perspectives. *Critical Reviews in Plant Sciences*, 25(5), 417–440. <https://doi.org/10.1080/07352680600899973>
- Dunse, K. M., Stevens, J. A., Lay, F. T., Gaspar, Y. M., Heath, R. L., & Anderson, M. A. (2010). Coexpression of potato type I and II proteinase inhibitors gives cotton plants protection against insect damage in the field. *Proceedings of the National Academy of Sciences*, 107(34), 15011–15015. <https://doi.org/10.1073/pnas.1009241107>
- Ehrlich, P. R., & Raven, P. H. (1964). Butterflies and Plants: A Study in Coevolution. *Evolution*, 18(4), 586–608. <https://doi.org/10.1111/j.1558-5646.1964.tb01674.x>
- Farley-Barnes, K. I., Ogawa, L. M., & Baserga, S. J. (2019). Ribosomopathies: Old Concepts, New Controversies. *Trends in Genetics*, 35(10), 754–767. <https://doi.org/10.1016/j.tig.2019.07.004>
- Fauteux, F., RÅmus-Borel, W., Menzies, J. G., & BÅlanger, R. R. (2005). Silicon and plant disease resistance against pathogenic fungi. *FEMS Microbiology Letters*, 249(1), 1–6. <https://doi.org/10.1016/j.femsle.2005.06.034>
- Ferlian, O., Thakur, M. P., Castañeda González, A., San Emeterio, L. M., Marr, S., da Silva Rocha, B., & Eisenhauer, N. (2020). Soil chemistry turned upside down: a meta-analysis of invasive earthworm effects on soil chemical properties. *Ecology*, 101(3), e02936. <https://doi.org/10.1002/ecy.2936>
- Figuroa-Macías, J. P., García, Y. C., Núñez, M., Díaz, K., Olea, A. F., & Espinoza, L. (2021). Plant Growth-Defense Trade-Offs: Molecular Processes Leading to Physiological Changes. *International Journal of Molecular Sciences*, 22(2), 693. <https://doi.org/10.3390/ijms22020693>
- Forrister, D. L. (2022). *Exploring the Phytochemical Landscape in Space and Time: Implications for the Ecology and Evolution of Tropical Trees and Their Insect Herbivores*.
- Frew, A., Weston, L. A., Reynolds, O. L., & Gurr, G. M. (2018). The role of silicon in plant biology: a paradigm shift in research approach. *Annals of Botany*, 121(7), 1265–1273. <https://doi.org/10.1093/aob/mcy009>
- Glas, J., Schimmel, B., Alba, J., Escobar-Bravo, R., Schuurink, R., & Kant, M. (2012). Plant Glandular Trichomes as Targets for Breeding or Engineering of Resistance to Herbivores. *International Journal of Molecular Sciences*, 13(12), 17077–17103. <https://doi.org/10.3390/ijms131217077>
- Glassmire, A. E., Jeffrey, C. S., Forister, M. L., Parchman, T. L., Nice, C. C., Jahner, J. P., Wilson, J. S., Walla, T. R., Richards, L. A., Smilanich, A. M., Leonard, M. D., Morrison, C. R., Simbaña, W., Salagaje, L. A., Dodson, C. D., Miller, J. S., Tepe, E. J., Villamarin-Cortez, S., & Dyer, L. A. (2016). Intraspecific phytochemical variation shapes community and population structure for specialist caterpillars. *New Phytologist*, 212(1), 208–219. <https://doi.org/10.1111/nph.14038>

- González-Sarrías, A., Espín, J. C., & Tomás-Barberán, F. A. (2017). Non-extractable polyphenols produce gut microbiota metabolites that persist in circulation and show anti-inflammatory and free radical-scavenging effects. *Trends in Food Science & Technology*, 69, 281–288. <https://doi.org/10.1016/j.tifs.2017.07.010>
- Greenslade, A. F. C., Ward, J. L., Martin, J. L., Corol, D. I., Clark, S. J., Smart, L. E., & Aradottir, G. I. (2016). Triticum monococcum lines with distinct metabolic phenotypes and phloem-based partial resistance to the bird cherry–oat aphid *Rhopalosiphum padi*. *Annals of Applied Biology*, 168(3), 435–449. <https://doi.org/10.1111/aab.12274>
- Griese, E. (2021). *Battle between insect eggs and host plants: Ecology and evolution of pierid egg-induced responses in Brassicaceae*.
- Hanley, M. E., Lamont, B. B., Fairbanks, M. M., & Rafferty, C. M. (2007). Plant structural traits and their role in anti-herbivore defence. *Perspectives in Plant Ecology, Evolution and Systematics*, 8(4), 157–178. <https://doi.org/10.1016/j.ppees.2007.01.001>
- Hartl, M., Giri, A. P., Kaur, H., & Baldwin, I. T. (2011). Serine Protease Inhibitors Specifically Defend *Solanum nigrum* against Generalist Herbivores but Do Not Influence Plant Growth and Development. *The Plant Cell*, 22(12), 4158–4175. <https://doi.org/10.1105/tpc.109.073395>
- He, J., Chen, F., Chen, S., Lv, G., Deng, Y., Fang, W., Liu, Z., Guan, Z., & He, C. (2011). Chrysanthemum leaf epidermal surface morphology and antioxidant and defense enzyme activity in response to aphid infestation. *Journal of Plant Physiology*, 168(7), 687–693. <https://doi.org/10.1016/j.jplph.2010.10.009>
- Hehmeyer, J. (2019). Two potential evolutionary origins of the fruiting bodies of the dictyostelid slime moulds. *Biological Reviews*, 94(5), 1591–1604. <https://doi.org/10.1111/brv.12516>
- Hollingsworth, R. G., Armstrong, J. W., & Campbell, E. (2002). Caffeine as a repellent for slugs and snails. *Nature*, 417(6892), 915–916. <https://doi.org/10.1038/417915a>
- Howe, G. A., & Jander, G. (2008). Plant Immunity to Insect Herbivores. *Annual Review of Plant Biology*, 59(1), 41–66. <https://doi.org/10.1146/annurev.arplant.59.032607.092825>
- Howe, G. A., & Jander, G. (2008). Plant immunity to insect herbivores. *Annu. Rev. Plant Biol.*, 59(1), 41–66. <https://doi.org/10.1146/annurev.arplant.59.032607.092825>
- Howe, G. A., & Schaller, A. (2008). Direct Defenses in Plants and Their Induction by Wounding and Insect Herbivores. *Induced Plant Resistance to Herbivory*, 7–29. https://doi.org/10.1007/978-1-4020-8182-8_1
- Hunter, J. E., Gannon, T. W., Richardson, R. J., Yelverton, F. H., & Leon, R. G. (2020). Integration of remote-weed mapping and an autonomous spraying unmanned aerial vehicle for site-specific weed management. *Pest Management Science*, 76(4), 1386–1392. <https://doi.org/10.1002/ps.5651>
- Islam, T., Moore, B. D., & Johnson, S. N. (2022). Plant silicon defences reduce the performance of a chewing insect herbivore which benefits a contemporaneous sap-feeding insect. *Ecological Entomology*, 47(6), 951–958. <https://doi.org/10.1111/een.13183>
- Jeblick, T., Leisen, T., Steidele, C. E., Albert, I., Müller, J., Kaiser, S., Mahler, F., Sommer, F., Keller, S., Hüchelhoven, R., Hahn, M., & Scheuring, D. (2023). Botrytis hypersensitive response inducing protein 1 triggers noncanonical PTI to induce plant cell death. *Plant Physiology*, 191(1), 125–141. <https://doi.org/10.1093/plphys/kiac476>
- Karban, R., & Baldwin, I. T. (2007). *Induced responses to herbivory*. University of Chicago Press.
- Keeping, M. G., Kvedaras, O. L., & Bruton, A. G. (2009). Epidermal silicon in sugarcane: Cultivar differences and role in resistance to sugarcane borer *Eldana saccharina*. *Environmental and Experimental Botany*, 66(1), 54–60. <https://doi.org/10.1016/j.envexpbot.2008.12.012>
- Kempema, L. A., Cui, X., Holzer, F. M., & Walling, L. L. (2007). Arabidopsis Transcriptome Changes in Response to Phloem-Feeding Silverleaf Whitefly Nymphs. Similarities and Distinctions in Responses to Aphids. *Plant Physiology*, 143(2), 849–865. <https://doi.org/10.1104/pp.106.090662>
- Leroy, N., de Tombeur, F., Walgraffe, Y., Cornélis, J.-T., & Verheggen, F. J. (2019). Silicon and Plant Natural Defenses against Insect Pests: Impact on Plant Volatile Organic Compounds and Cascade Effects on Multitrophic Interactions. *Plants*, 8(11), 444. <https://doi.org/10.3390/plants8110444>
- Levin, D. A. (1973). The Role of Trichomes in Plant Defense. *The Quarterly Review of Biology*, 48(1, Part 1), 3–15. <https://doi.org/10.1086/407484>
- Lev-Yadun, S. (2016). Ant Mimicry. *Defensive (Anti-Herbivory) Coloration in Land Plants*, 299–304. https://doi.org/10.1007/978-3-319-42096-7_58
- Liang, H.-J., Di, Y.-L., Li, J.-L., & Zhu, F.-X. (2015). Baseline sensitivity and control efficacy of fluazinam against *Sclerotinia sclerotiorum*. *European Journal of Plant Pathology*, 142(4), 691–699. <https://doi.org/10.1007/s10658-015-0644-5>

- Lindenmayer, D. B., & Franklin, J. F. (2013). *Conserving forest biodiversity: a comprehensive multiscaled approach*.
- Lugtenberg, B. J. J., Caradus, J. R., & Johnson, L. J. (2016). Fungal endophytes for sustainable crop production. *FEMS Microbiology Ecology*, 92(12), fiw194. <https://doi.org/10.1093/femsec/fiw194>
- Ma, H.-Y., Li, Y.-Y., Li, L., Tan, Y., & Pang, B.-P. (2021). Regulation of Juvenile Hormone on Summer Diapause of *Geleruca daurica* and Its Pathway Analysis. *Insects*, 12(3), 237. <https://doi.org/10.3390/insects12030237>
- Ma, J. F., & Yamaji, N. (2006). Silicon uptake and accumulation in higher plants. *Trends in Plant Science*, 11(8), 392–397. <https://doi.org/10.1016/j.tplants.2006.06.007>
- Maffei, M. E. (2010). Sites of synthesis, biochemistry and functional role of plant volatiles. *South African Journal of Botany*, 76(4), 612–631. <https://doi.org/10.1016/j.sajb.2010.03.003>
- Massey, F. P., Ennos, A. R., & Hartley, S. E. (2006). Silica in grasses as a defence against insect herbivores: contrasting effects on folivores and a phloem feeder. *Journal of Animal Ecology*, 75(2), 595–603. <https://doi.org/10.1111/j.1365-2656.2006.01082.x>
- Matsui, K., & Engelberth, J. (2022). Green Leaf Volatiles the Forefront of Plant Responses Against Biotic Attack. *Plant and Cell Physiology*, 63(10), 1378–1390. <https://doi.org/10.1093/pcp/pcac117>
- Middleton, E., Kandaswami, C., & Theoharides, T. C. (2000). The effects of plant flavonoids on mammalian cells: Implications for inflammation, heart disease, and cancer. *Pharmacological Reviews*, 52(4), 673–751.
- Miettinen, K., Dong, L., Navrot, N., Schneider, T., Burlat, V., Pollier, J., Woittiez, L., van der Krol, S., Lugin, R., Ilc, T., Verpoorte, R., Oksman-Caldentey, K.-M., Martinoia, E., Bouwmeester, H., Goossens, A., Memelink, J., & Werck-Reichhart, D. (2014). The seco-iridoid pathway from *Catharanthus roseus*. *Nature Communications*, 5(1), 3606. <https://doi.org/10.1038/ncomms4606>
- Musaqaf, N., Lyngs Jørgensen, H. J., & Sigsgaard, L. (2023). Plant resistance induced by hemipterans Effects on insect herbivores and pathogens. *Crop Protection*, 163, 106122. <https://doi.org/10.1016/j.cropro.2022.106122>
- Naalden, D., van Kleeff, P. J. M., Dangol, S., Mastop, M., Corkill, R., Hogenhout, S. A., Kant, M. R., & Schuurink, R. C. (2021). Spotlight on the Roles of Whitefly Effectors in Insect–Plant Interactions. *Frontiers in Plant Science*, 12, 661141. <https://doi.org/10.3389/fpls.2021.661141>
- Nallu, S., Hill, J. A., Don, K., Sahagun, C., Zhang, W., Meslin, C., Snell-Rood, E., Clark, N. L., Morehouse, N. I., Bergelson, J., Wheat, C. W., & Kronforst, M. R. (2018). The molecular genetic basis of herbivory between butterflies and their host plants. *Nature Ecology & Evolution*, 2(9), 1418–1427. <https://doi.org/10.1038/s41559-018-0629-9>
- Paljakka, T. (2020). Tree water transport mediating the changing environmental conditions to tree physiological processes. *Dissertationes Forestales*, 2020(302). <https://doi.org/10.14214/df.302>
- Pavela, R. (2016). History, presence and perspective of using plant extracts as commercial botanical insecticides and farm products for protection against insects - a review. *Plant Protection Science*, 52(4), 229–241. <https://doi.org/10.17221/31/2016-pps>
- Petschenka, G., & Agrawal, A. A. (2016). How herbivores coopt plant defenses: natural selection, specialization, and sequestration. *Current Opinion in Insect Science*, 14, 17–24. <https://doi.org/10.1016/j.cois.2015.12.004>
- Rattan, R. S. (2010). Mechanism of action of insecticidal secondary metabolites of plant origin. *Crop Protection*, 29(9), 913–920. <https://doi.org/10.1016/j.cropro.2010.05.008>
- Raubenheimer, D., Simpson, S. J., & Mayntz, D. (2009). Nutrition, ecology and nutritional ecology: toward an integrated framework. *Functional Ecology*, 23(1), 4–16. <https://doi.org/10.1111/j.1365-2435.2009.01522.x>
- Raupp, M. J. (1985). Effects of leaf toughness on mandibular wear of the leaf beetle, *Plagioderma versicolora*. *Ecological Entomology*, 10(1), 73–79. <https://doi.org/10.1111/j.1365-2311.1985.tb00536.x>
- Reynolds, O. L., Padula, M. P., Zeng, R., & Gurr, G. M. (2016). Silicon: Potential to Promote Direct and Indirect Effects on Plant Defense Against Arthropod Pests in Agriculture. *Frontiers in Plant Science*, 7, 744. <https://doi.org/10.3389/fpls.2016.00744>
- Rosenhek-Goldian, I., Kampf, N., Yeredor, A., & Klein, J. (2015). On the question of whether lubricants fluidize in stick–slip friction. *Proceedings of the National Academy of Sciences*, 112(23), 7117–7122. <https://doi.org/10.1073/pnas.1505609112>
- Schoonhoven, L. M., Loon, J. J., & Dicke, M. (2005). *Insect-plant biology*.
- Schulze, E.-D., Beck, E., Buchmann, N., Clemens, S., Müller-Hohenstein, K., & Scherer-Lorenzen, M. (2019). Interactions Between Plants, Plant Communities and the Abiotic and Biotic Environment. *Plant Ecology*, 689–741. https://doi.org/10.1007/978-3-662-56233-8_19

- Seal, P., Das, P., & Biswas, A. K. (2018). Versatile Potentiality of Silicon in Mitigation of Biotic and Abiotic Stresses in Plants: A Review. *American Journal of Plant Sciences*, 09(07), 1433–1454. <https://doi.org/10.4236/ajps.2018.97105>
- Shivaji, R., Camas, A., Ankala, A., Engelberth, J., Tumlinson, J. H., Williams, W. P., Wilkinson, J. R., & Luthe, D. S. (2010). Plants on Constant Alert: Elevated Levels of Jasmonic Acid and Jasmonate-Induced Transcripts in Caterpillar-Resistant Maize. *Journal of Chemical Ecology*, 36(2), 179–191. <https://doi.org/10.1007/s10886-010-9752-z>
- Silva, W. T. A. F., Hansson, M., & Johansson, J. (2021). Light competition and phenological adaptation of annual plants to a changing climate. *Climate Change Ecology*, 2, 100007. <https://doi.org/10.1016/j.ecochg.2021.100007>
- Silva-Junior, N. R., Meriño Cabrera, Y. B., de Almeida Barros, R., & de Almeida Oliveira, M. G. (2022). Use of Protease Inhibitors as a Promising Alternative for Pest Control. *Natural Products as Enzyme Inhibitors: An Industrial Perspective*, 137–151. https://doi.org/10.1007/978-981-19-0932-0_6
- Singh, B., & Kaur, A. (2018). Control of insect pests in crop plants and stored food grains using plant saponins: A review. *LWT*, 87, 93–101. <https://doi.org/10.1016/j.lwt.2017.08.077>
- Song, Y., & Thomma, B. P. H. J. (2018). Host-induced gene silencing compromises Verticillium wilt in tomato and *Arabidopsis*. *Molecular Plant Pathology*, 19(1), 77–89. <https://doi.org/10.1111/mpp.12500>
- Steppuhn, A., & Baldwin, I. T. (2007). Resistance management in a native plant: nicotine prevents herbivores from compensating for plant protease inhibitors. *Ecology Letters*, 10(6), 499–511. <https://doi.org/10.1111/j.1461-0248.2007.01045.x>
- Tan, C.-W. (2019). *Indirect Manipulation of Plant Induced Defenses by Parasitoids of Caterpillars*.
- Tscharntke, T., Thiessen, S., Dolch, R., & Boland, W. (2001). Herbivory, induced resistance, and interplant signal transfer in *Alnus glutinosa*. *Biochemical Systematics and Ecology*, 29(10), 1025–1047. [https://doi.org/10.1016/s0305-1978\(01\)00048-5](https://doi.org/10.1016/s0305-1978(01)00048-5)
- Turlings, T. C. J., & Erb, M. (2018). Tritrophic Interactions Mediated by Herbivore-Induced Plant Volatiles: Mechanisms, Ecological Relevance, and Application Potential. *Annual Review of Entomology*, 63(1), 433–452. <https://doi.org/10.1146/annurev-ento-020117-043507>
- Usha Rani, P., & Jyothsna, Y. (2010). Biochemical and enzymatic changes in rice plants as a mechanism of defense. *Acta Physiologiae Plantarum*, 32(4), 695–701. <https://doi.org/10.1007/s11738-009-0449-2>
- van Loon, L. C., Geraats, B. P. J., & Linthorst, H. J. M. (2006). Ethylene as a modulator of disease resistance in plants. *Trends in Plant Science*, 11(4), 184–191. <https://doi.org/10.1016/j.tplants.2006.02.005>
- Vicente, M. R.-S., & Plasencia, J. (2011). Salicylic acid beyond defence: its role in plant growth and development. *Journal of Experimental Botany*, 62(10), 3321–3338. <https://doi.org/10.1093/jxb/err031>
- War, A. R., Paulraj, M. G., War, M. Y., & Ignacimuthu, S. (2011). Jasmonic Acid-Mediated-Induced Resistance in Groundnut (*Arachis hypogaea* L.) Against *Helicoverpa armigera* (Hubner) (Lepidoptera: Noctuidae). *Journal of Plant Growth Regulation*, 30(4), 512–523. <https://doi.org/10.1007/s00344-011-9213-0>
- Watson, G. W., Adalla, C. B., Shepard, B. M., & Carner, G. R. (2015). *Aspidiotus rigidus* (Hemiptera: Diaspididae): a devastating pest of coconut in the Philippines. *Agricultural and Forest Entomology*, 17(1), 1–8. <https://doi.org/10.1111/afe.12074>
- Whitehill, J. G. A., Bohlmann, J., & Krokene, P. (2023). Forest Insect—Plant Interactions. *Forest Entomology and Pathology: Volume 1: Entomology*, 169–204. https://doi.org/10.1007/978-3-031-11553-0_7
- Yang, M., Liu, C., Zhao, T., Li, Y., Liu, H., Ren, Z., & Xue, K. (2023). Trichomes on Cotton Leaf Surface Affect the Feeding Behaviors of Cotton Aphids. *Research Square*. <https://doi.org/10.21203/rs.3.rs-2476798/v1>
- Zargar, S. M., Mahajan, R., Bhat, J. A., Nazir, M., & Deshmukh, R. (2019). Role of silicon in plant stress tolerance: Opportunities to achieve a sustainable cropping system. *Biotech*, 9(3), 73. <https://doi.org/10.1007/s13205-019-1613-z>
- Züst, T., & Agrawal, A. A. (2017). Trade-offs between plant growth and defense against insect herbivory: An emerging mechanistic synthesis. *Annual Review of Plant Biology*, 68(1), 513–534. <https://doi.org/10.1146/annurev-arplant-042916-040856>